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Muscle, Tendon, and Biomechanics

A Subset of FAP Cells Expressing Gli1 Promote Muscle Regeneration With Less Fat Accumulation

Disclosures

None

Introduction

Skeletal muscle has a remarkable capacity for regeneration after injury. Recently, a new type of muscle-resident progenitor cell, referred to as fibro-adipogenic progenitors (FAPs), was identified to be critical in supporting the process of injured muscle regeneration. To date, FAPs remain a poorly defined, heterogeneous population without any specific genetic markers. Using a lineage tracing mouse model (Gli1-CreER Tomato, Gli1ER/Td), we recently discovered that Gli1 marks a small subset of muscle-resident FAPs (17%) that preferentially expand upon muscle injury (40% of FAPs at day 3 after injury). Here, we performed cell ablation, pharmacologic manipulation, and single cell transcriptomics to further investigate the role of Gli1⁺ FAPs in muscle regeneration and fat

Methods

Animals

All animal work was approved by the Institutional Animal Care and Use Committee (IACUC) at the University of Pennsylvania. *Gli1-CreER Rosa-tdTomato (Gli1ER/Td)* mice and *Gli1-CreER Rosa-tdTomato DTA (Gli1ER/Td/DTA)* mice were generated. To induce *CreER* activity, 2-month-old male mice received tamoxifen (Tam) injections (75 mg/kg/day) for 5 days. Acute muscle injury was induced by injection of 10 μ l Notexin (NTX, 10 μ g/mL) or 50 μ l glycerol (50% vol/vol) into the Tibialis Anterior (TA) muscle.

Histology

TA muscles were processed for cryosections followed by H&E, WGA, or Perilipin antibodies staining.

Single cell RNA sequencing (scRNA-seq) analysis

Pre-aligned and filtered single-cell RNA-seq matrix files of mouse muscle cells were acquired from GEO: GSE110878. Unsupervised clustering

was conducted by Seurat and GO/KEGG term enrichment were analyzed by Clusterprofiler.

Statistics

Data are expressed as means \pm standard error of the mean and analyzed by t-tests or one-way ANOVA.

Results

In the TA muscle of *Gli1ER/Td* mice, Td⁺ cells were exclusively FAPs located in the interstitial area of myofibers (Figure 1A). Though initially presented at a low level in freshly digested muscle cells, Td⁺ cells constituted 40% of confluent cells after culturing. Sorted Td⁺ FAPs formed 6.2-

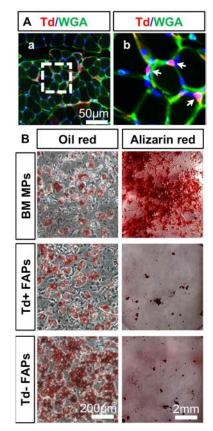


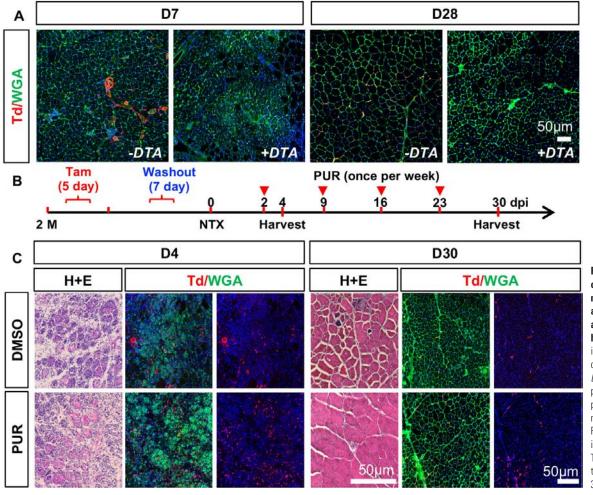
Figure 1. Td in the muscle of *Gli1ER/Td* mice labels a subpopulation of FAPs with less adipogenicity. (A) Td^+ cells located in the interstitial area of TA muscle in *Gli1ER/Td* mice. (B) Representative images of osteogenic and adipogenic differentiation of BM MPs (Bone marrow mesenchymal progenitors), Td^+ FAPs and Td FAPs.

fold more CFU-Fs than Td⁻ FAPs. They had fibroblastic and adipogenic differentiation abilities, but generated much fewer adipocytes than Td⁻ FAPs (Figure 1B). Both Td⁺ and Td⁻ cells had no osteogenic differentiation ability (Figure 1B). To investigate their *in vivo* function, we generated *Gli1ER/Td/ DTA* mice. At 2 months of age, Tam injections quickly reduced Td⁺ cells by 80% and eliminated Td⁺ CFU-F colonies from muscle (n = 3/group), indicating successful cell ablation. Interestingly, myofibers regenerated at day 7 and day 28 in *Gli1ER/Td/DTA* mice after Notexin injury were 34.1% and 22.7%, respectively. These were smaller than those in *Gli1ER/Td Td* mice (Figure 2A, n = 5/group), suggesting an impairment of myogenesis.

Gli1 is an effector of Hedgehog (Hh) signaling. Intramuscular injection of purmorphamine (PUR), an Hh agonist, expanded Td⁺ cells at day 4 and increased myofiber size by 1.6-fold at day 30 after Notexin injury (Figure 2B, C, n = 4/group). Local glycerol injection causes muscle degeneration with fat infiltration. Strikingly, Td⁺ FAPs were less likely to become adipocytes compared to Td⁻ FAPs and ablation of Td⁺ cells led to 39.9% more adipocytes in muscle (Figure 3A-D, n = 4/group). Hh signaling inhibitor, GANT61, induced 78.3% more adipocytes, while PUR almost completely depleted adipocyte infiltration (n = 5/group). ScRNA-seq analysis of mouse muscle mononucleated cells showed that FAPs can be subclustered into Gli1⁺ and Gli1⁻ FAPs (Figure 3E, F). Pathway analysis suggested that Gli1⁺ FAPs are more metabolic active and more related to tissue regeneration than Gli1⁻ FAPs. For example, they have higher Interleukin-6 (II6) production and TGF-beta signaling, two known positive regulators of myoblast proliferation and tissue regeneration. Furthermore, Gli1⁺ FAPs express higher levels of known muscle regulators, such as Tgfb1, Wisp1, Malat1, Igf1, Il15 and Il33 (Figure 3G), compared with Gli1⁻ FAPs, which was further validated by qRT-PCR (Figure 3H). Il15 and *Wisp1*, are also involved in inhibiting FAPs differentiation into adipocytes. Further analysis revealed increased expression of anti-adipogenic regulators, such as Tsc22d3, Dlk1, Ddit3 and Nr4a1 (Figure 3I), and reduced expression of pro-adipogenic regulator, such as Zfp423 and Ebf1 (Figure 3J).

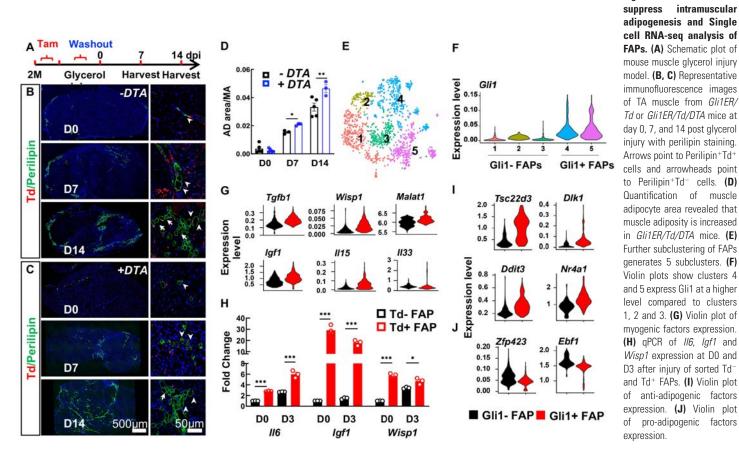
Discussion

Our study found that *Gli1-CreER* labels a subpopulation of FAP cells. Compared to Gli1⁻ FAPs, Gli1⁺ FAPs are more metabolically active for muscle repair and less likely to contribute to muscle adiposity. Our studies demonstrated that Hh/Gli1 signaling pathway is critical for regulating muscle regeneration and fat accumulation, indicating that



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Figure 2. Ablation of Gli1+ cells delayed causes muscle regeneration and activation of Hh signaling accelerates muscle healing. (A) Representative immunofluorescence imaging of Gli1ER/Td and Gli1ER/Td/ DTA muscle at day 7 and 28 post NTX injury. (B) Schematic plot of mouse muscle NTX injury model with PUR treatment. (C) Representative H&E staining and immunofluorescence images of TA muscles with DMSO or PUR treatment group at day 4 and day 30 post NTX injury.



pharmacological activation of this pathway could be a therapeutic approach to boost muscle regeneration.

accumulation and implied a potential therapeutic effect of Hedgehog signaling in muscle diseases.

Significance

Our study revealed a subpopulation of FAPs that preferentially promotes muscle regeneration with less fat

FAPs

Figure 3. Gli1⁺